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Mechanisms of Drought Tolerance in Sorghum: A Review

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ABSTRACT

Sorghum is the fifth most important cereal crop and occupies the second position among the staple food grains in semi-arid tropics. The adaptation of grain sorghum to a wide range of environmental conditions has led to the evolution and existence of extensive genetic variation for drought tolerance. Accordingly, sorghum is expected to play an increasingly important role in agriculture and meeting world food demand in the face of climate change, land degradation and increasing water scarcity. The crop requires relatively less water than other important cereals such as maize and wheat. However, yield potential of the crop is significantly limited due to drought and heat stresses. Drought is one of the most important factors that affect crop production worldwide and continues to be a challenge to plant breeders, despite many decades of research. Underestimating the different mechanisms underlying drought tolerance is vital for the breeding to alleviate adverse effects of drought in order to boost productivity. In this literature review, the main effects of drought on crop growth and development, and yield are reported.

Key words: Drought, Sorghum, Physiological mechanisms, Drought tolerance, Breeding.

INTRODUCTION

Drought stress is a serious agronomic problem contributing to severe yield losses worldwide. This agricultural constraint may nevertheless be addressed by developing crops that are well adapted to drought prone environments. Drought tolerance depends on the plant developmental stage at the onset of the stress syndrome, which in sorghum may happen during the early vegetative seedling stage, during panicle development and in postflowering, in the period between grain filling and physiological maturity 93,94.

Sorghum is one of the most drought tolerant crop species and is an important model system for studying physiological and molecular mechanisms underlying drought tolerance. It is considered as one of the most important crops for production cereal grains and fodder for humans and animals. Also grain yield used to bridge the deficit in wheat flour to produce bread municipal.

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A biotic stress such as water stress and salinity was the incumbent in the first resistor work to improve the resistance for environmental stresses, especially the problem of water shortage. Sorghum is predictable to play an increasingly significantly function in cultivation and gathering world food desire in the countenance of climate modification, ground degradation and mounting water reduction⁴.

Sorghum serves as a dietary staple crop for millions of people, especially in arid and semi-arid farming systems. Additionally, sorghum grain is used as livestock feed and for production of local beverages, while the stalk is used for animal feed, firewood, and as a construction material⁶⁸. Sorghum grows across a wide geographic area at various altitude, day length, rainfall, and temperature regimes. Consequently, it is well adapted to withstand harsh conditions, which are the characteristic feature of tropical regions. The crop requires relatively less water than other important cereals such as maize and wheat. However, yield potential of the crop is significantly limited due to drought and heat stresses within the tropics and subtropics necessitating sorghum breeding for drought tolerance and productivity¹⁴.

Drought can occur at any stages of the crop development. However, in the arid and semi-arid tropics, the probability of drought is highest at the start and end of the growing season. Drought stress at the beginning of the growing season will severely affect plant establishment. If drought occurs at flowering, or in the grain filling stages, it may result in reduced yield, or complete crop failure ¹².

Drought contributes to poor crop performance and yield. Countries in arid and semi-arid tropics usually experience insufficient, unevenly distributed, unpredictable rainfall. At one point rain may be abundant and perhaps wasted through runoff; in some years much rain may fall completely outside the growing season. In other years, in adequate mid-season rain may fall after crops have germinated, causing crop failure. Although drought stress at the

beginning of the growing season may severely affect plant establishment, plants tend to recover soon when late rain fall levels are adequate⁸⁵. Consequently, crops are prone to periodic moisture stress in one way or another because of the aforementioned realities 116. The impact of moisture stress on crop yield is dependent the stage on of development^{53,111}. Anthesis and grain filling stages appear to be the most vulnerable growth stages; occurrence of drought at these stages may result in reduced yield and/or complete crop failure¹²⁸.

Crop production is constrained by several biotic, abiotic and socio-economic factors. Amongst the most important abiotic constraints, drought is the most important. Therefore, understanding of the physiological mechanisms and genetic control of drought in crops is important as a base for improving the production and productivity of crops in the arid and semi-arid tropics. In this article, different mechanism of drought tolerance in crops have been reviewed.

Effects of Drought on Crop Growth and Development

Drought is a combination of stress effects caused by high temperatures⁸⁰ and a lack of water²². Evapo-transpiration is the major driving force that affects the soil, plant, and atmospheric continuum of the hydrologic cycle. In earlier studies, predictions of drought were mainly based on the amount and distribution of precipitation¹³. However, in recent studies soil moisture balance and soil characteristics have been introduced in the assessment of drought. Lack of adequate soil moisture, or water deficit, affects the ability of plants to grow and complete a normal life cycle⁷⁴.

Drought can have major consequences on growth, development and yield of crops by affecting several physiological, morphological and biochemical processes¹⁰³. It is the major cause of poor crop performance and low yields, and sometimes it causes total crop failure. In the tropics, the probability of drought is highest at the start and the end of the growing season.

Drought can occur at both seedling, preflowering and post-flowering stages of development, and has the most adverse effect on yield^{53,111}. Drought stress at the seedling stage of development will severely affect plant establishment⁵. If it occurs at flowering, or in the grain filling stages, it may cause reduced yields, or complete crop failure¹². Researchers have classified drought as either pre- or postflowering stress. The reactions of genotypes to these stresses are variable and controlled by different genetic mechanisms. Pre-anthesis moisture stress has effects on components such as stand count, tillering capacity, number of heads and number of seeds per head, while post-anthesis moisture stress affects transpiration efficiency, CO2 fixation and carbohydrate translocation. The latter factors, in turn, results in low yields and premature plant senescence 109,123.

Physiological Mechanisms of Drought Tolerance

Levitt⁵⁸ mentioned that drought resistance mechanisms like drought avoidance, recovery, survival and tolerance, are associated with plant survival and production. Drought avoidance is defined as the ability of plants to conserve water at the whole plant level through decreasing water loss from the shoots or by more efficiently extracting water from the soil⁶⁰. However, drought tolerance is defined as the ability of plants to withstand water deficit while maintaining appropriate physiological activities to stabilize and protect cellular and metabolic integrity at tissue and cellular level^{111,124}. Survival is the ability of the crop to survive drought, irrespective of the yield it produces, while production is the ability of the crop to grow and yield under water stress conditions⁴.

Ceccarelli *et al*²³., defined drought tolerance, is a complex quantitative trait influenced by many genetic and environmental factors. The responses of different plants, species, or genotypes to drought are variable in relation to developmental stage, duration of drought, and evolutionary adaptation of the crop⁹⁷. In sorghum, for example, varieties that are adapted to arid and semi-arid environments

showed higher drought tolerance than varieties of humid origin⁹. Several studies have been conducted in understanding the mechanism of drought resistance in crops and in identifying essential traits for drought tolerance¹³. Drought resistance, therefore, involves the interaction of different morphological structures, physiological functions, and biochemical expressions^{20,71}.

Stomatal Conductance and Leaf Rolling

In plants, stomatal conductance and leaf rolling have been found to be reliable physiological indicators of drought tolerance⁵², which are strongly associated with leaf water potential⁷. These two mechanisms controlled by different factors, where stomatal conductance is controlled by soil moisture dependent root signals, while leaf rolling is controlled by leaf water potential²⁸. The strong correlation of leaf rolling and leaf water potential allows breeders to use leaf rolling as a visual scoring criterion for selecting for drought resistance in plants⁴⁶. The rolling of leaves usually occurs following the reduction in leaf water potential. However, the degree of leaf rolling depends on the ability of the plant to adjust osmotically at low leaf water potential³⁵. Plants with high osmotic adjustment develop less leaf rolling, and hence, reduced leaf rolling is considered as an indicator of a greater degree of desiccation avoidance, through a deep root system⁴⁶.

Drought tolerant genotypes exhibit lower stomatal conductance associated with increased leaf temperature, which gives rise to high transpiration efficiency and lower carbon isotope discrimination. The drought susceptible genotypes, on the other hand, show higher stomatal conductance and lower leaf temperature results in lower transpiration rates⁵⁴.

There is a lack of consensus on the benefits of the two traits of leaf rolling and stomatal conductance as drought resistance mechanisms. Leaf rolling has a detrimental effect on transpiration rate through changes in leaf stomatal conductance, and reduction in effective leaf area⁸⁸. In addition, leaf rolling enhances stomatal closure by increasing leaf

resistance to water loss. However, Heckathorn and DeLucia⁴³ argued that leaf rolling had positive effects on reducing leaf temperature and loss of water by decreasing the incident irradiation. Stomatal closure alone causes a 70-80 % decrease in transpiration rate in crops, where leaf rolling causes a decrease of only 2 % of normal transpiration rate⁴³. Therefore, leaf rolling has less value in reducing water loss than stomatal closure and it may increase the survival of plants by enhancing stomatal closure under extreme drought conditions⁴³. The significance of using these traits as physiological indicators of plant drought adaptive mechanisms depends on the crop species and the environment. Under conditions where there are no sophisticated instruments to measure transpiration efficiency and stomatal conductance, leaf rolling is good indicator of drought tolerance.

Characteristics of Root

Roots are the primary plant organ affected by drought stress and other environmental stresses of the soil⁸¹. Sorghum crown roots grow about 2 to 3 cm per day⁹⁶ and root growth is mainly affected by the amount of carbon partitioned to the roots, although it varies with environmental and genetic factors¹⁰. Sorghum roots may grow to depths of 1 to 2 m by the booting stage, and can efficiently extract water to a lateral distance of 1.6 m from the plant⁹⁶. Root growth in sorghum terminates at flowering stage; however, it is more prominent in a senescent than in nonsenescent sorghum genotypes⁹¹.

Bawazir and Idle⁶ reported variation in root anatomy and morphology, among sorghum genotypes. Genotypes that have large number of seminal roots, large vessel diameter in both seminal and nodal roots showed better survival rate under drought stress conditions. Similarly, Habyarimana *et al*³⁹., found that the drought tolerance traits displayed by the genotypes were related to drought avoidance mechanisms. These, in turn, are associated with deep root system, which enables plants to exploit moisture from the deeper soil horizons.

The root has received less attention than the shoot in the search for characters of

use for screening or selection for drought resistance. Esau³² remarked that in dry soil the restriction of adventitious root growth makes the efficiency of water transport depend more upon the conductivity of the seminal roots. Camacho *et al*²¹., showed that plants with efficient water transport systems avoid dehydration of the leaf tissue during periods of atmospheric drought.

Meyer and Ritchie⁷⁰ showed that, the root contributes more resistance than the shoot at least at high transpiration rates. Richards and Passioura⁸⁹ demonstrated variation in vessel size related to climatic factors in some wheat accessions, and proposed a selection and breeding programme for small vessels in the expectation of improving the performance of wheat under conditions of limited water supply. Passioura⁷⁸ reports encouraging signs in this programme.

Ekanayake et al^{31} ., indicated that drought stress tolerance was found to be highly associated with root characteristics such as root thickness, root length density, number of thick roots, root volume, and root dry weight. It was also found that number of thick root, root thickness, and root length density were highly associated with leaf water potential and field visual drought scoring using drying leaf. Drought stress adapted plants are often characterized by deep and vigorous root systems¹⁵. Nour et al⁷⁶., also reported root weight is the best and easiest attribute to determine drought tolerance in grain sorghum. Matsuura et al^{67} ., on the other hand, reported a positive correlation between drought tolerance and root length in sorghum and millet (Pennisetum glaucum). Moreover, Plaut et al^{79} ., and Pace et al^{77} ., reported that seedlings under water stress caused an increase in root length with reduced diameter. Root depth, root length density, root distribution was reported as drought tolerance contributing traits 106.

Drought is often associated with nutrient availability and the capacity of roots to absorb the available nutrients. Ludlow and Muchow⁶⁰ indicated that greater root activity under intermittent drought should enhance crop stability by reducing the incidence of

water deficits. Egilla et al³⁰., and HongBo et al⁴⁵., reported the significance of potassium (K). in improving drought resistance and root longevity. Shao et al¹⁰¹., also reported the importance of mineral elements, such as K+ and Na⁺ for root signal transduction function. Shangguan et al⁹⁸., further denoted that the hydraulic conductivity of roots can be mainly affected by nitrogen and phosphorous nutrients. Hydraulic conductance in sorghum is primarily dependent on the number of fully functional nodal roots¹¹. In moisture stress conditions, plants with sufficient P supply exhibited higher hydraulic conductivity than P deficient plants. Therefore, plants with sufficient P are found to be more droughts tolerant, and also have a higher ability to recover after drought.

Osmotic Adjustment

Under rainfed conditions, plants are exposed to varying degrees of water stress due to lack of an adequate water supply to meet the transpirational demand. The ability of a crop to grow in areas subject to water deficits has been termed drought resistance 115. Individual plant species differ markedly in the mechanisms utilized to survive when water deficit exist. Adaptations to survive drought may be anatomical, morphological or physiological in nature and they serve either to faciliate the maintenance of favourable water balance (increasing water absorption or decreasing water loss). or to allow desiccation tolerance at low leaf ψ_w .

Osmotic adjustment is a major drought adaptive mechanism in plants⁴⁹. Sorghum and millet landraces, which are collected either dry or humid environments show variation in osmotic adjustment. Landraces that come from drier regions show greater osmotic adjustment than landraces from humid regions. The assumption is that through the course of evolution the drier environments provided sufficient selection pressure for osmotic adjustment. Landraces with higher osmotic adjustment are characterized by their dwarf nature with high rates of transpiration and low rates of leaf senescence under stress⁹.

adjustment improves Osmotic crop productivity through delaying leaf rolling and leaf tissue death¹³. As leaf rolling and leaf senescence decreases, the effective leaf area for photosynthesis increases. In a study by Ludlow et al⁶³., on sorghum genotypes, those with high osmotic adjustment exhibited a 24 % higher yield than genotypes with low adjustment, when exposed to a post-anthesis drought stress. The yield difference observed was both in grain size and grain number, and it was associated with higher harvest index. Similarly, Amede and Schubert³ observed that, a 20 % dry matter yield was increased in legume species that maintained turgor through osmotic adjustment. The contribution of osmotic adjustment to reducing yield losses varies with the intensity and duration of the stress⁴⁶. In general, yield reduction of stressed plants compared with non-stressed plants is due to the plant's additional energy requirements for osmotic adjustment 14,71.

Evidence for maintenance of stomatal conductance by turgor maintenance due to osmotic adjustment has been reported for field crops 1,61,62,113,114,122. Nevertheless, not all data have provided such confirmation. Jones and Rawson⁵⁰, Turner et al¹¹³., and Gollan et al³⁷., working with several species observed a range stomatal conductance and photosynthesis at zero turgor. Because of contradictory and limited information on the association of physiological parameters with osmotic adjustment, the question whether osmotic adjustment is beneficial in contributing to productivity or survival mechanism under water stress conditions is not yet answered.

Transpiration Efficiency

Transpiration efficiency (TE). is defined by Xin *et al*¹²³., it is a biomass accumulation per unit water transpired. Variation in TE within species has been demonstrated for several C3 plant species such as wheat, barley, rice, cotton, beans, tomato, and sunflower^{47,57,69,87,105,108}. Genetic variation in TE has also been found in sorghum using gas-exchange properties, traditional lysimetric assays, and field evaluation^{40,73}. Sorghum

genotypes with low internal CO_2 concentration and enhanced photosynthetic capacity may be associated with high TE, whereas high TE was strongly correlated with increased biomass accumulation, rather than with reduced water use¹²³.

Accumulation of Solutes

Solutes are low-molecular-weight and highly soluble compounds that are usually nontoxic high cytosolic concentrations. Generally, they protect plants from stress through different means such as contribution towards osmotic adjustment, detoxification of reactive oxygen species, stabilization of membranes, and native structures of enzymes and proteins³³. In sorghum, proline¹⁰⁴, glycinebetaine (GB). 127 and sugars functions as protect osmolytes that cells dehydration¹²⁰. GB accumulation in cells can assist plants to either maintain water within cells or protect cellular component from dehydration¹²⁷. However, the genetic and metabolic basis of variation GB accumulation is not well understood in sorghum²⁰. Grote et al³⁸., reported that a recessive allele of a single locus is associated with non-accumulation of GB in sorghum genotypes.

Accumulation of free proline in waterstressed sorghum leaves is related to the ability of a cultivar to recover from stress, possibly due to proline's role as a source of respiratory energy in the recovering plant⁸. In wheat, accumulation of proline⁹⁹ and anti-oxidative enzymes has been reported in both wild and cultivated species. Different wheat genotypes have different visible water threshold levels resulting in diverse responses to drought in terms of proline and anti-oxidative enzyme accumulation 100. Proline comprises 18 % of the osmotic pool in chickpea³. A strong accumulation of proline increases the cell solute concentration, resulting in increased water potential in the tissue through osmotic adjustment. Alternatively, the expression of anti-oxidative enzymes serves as a signal transduction for gene expression, and hence, proteins are synthesized, which control metabolism effluxes. Evaluation of rice

genotypes under *in vitro* drought induced conditions revealed a significant accumulation of proline and total soluble sugars in the leaves¹¹⁷. The tolerant lines showed a continuous increase in proline level for five weeks after the stress was induced and started to decline after six weeks under drought. The solute concentrations decreased to normal levels when plants were allowed to recover from drought stress¹¹⁷.

Grain formation and development in crop plants is dependent on assimilates produced by photosynthesis after anthesis or assimilates stored mainly in the stem before anthesis. Wheat genotypes revealed genotypic variation in the relative importance of preassimilates and post-anthesis anthesis photosynthesis to drought resistance⁴⁸. A relatively high photosynthetic rate during grain filling under water stress was observed in resistant cultivars relative susceptible cultivars. Moreover, the drought susceptible cultivars were much more reliant on remobilization of pre-anthesis assimilates stored in the stem to fill the grain as opposed to the resistant cultivar⁴⁸. This demonstrates that, under moisture stress, the pre-anthesis assimilates stored in the stem in the drought resistance cultivars are used to maintain a higher photosynthetic rate during the grain filling period.

Remobilization of pre-anthesis assimilates from the leaf and stem is one of the drought escape mechanism. In conditions where photosynthesis is inhibited by stress such as drought, heat, leaf diseases or shading, the demand for nutrient storage usually exists¹⁵. A large yield sink produces a physiological load on the leaves and stem, and the impact of this load is intensified under drought stress when the demand for carbon from stored reserves increases⁵⁵. However, Blum et al16., indicated that there are cases where the utilization of stem reserves for grain filling is not dictated by the environmental conditions. Genetically, male sterile plants showed a twofold increase in assimilate storage in the stems, indicating that removal of a grain sink increases stem sugar³⁶. Leaf

defoliation during anthesis promoted lodging, suggesting that it may be due to depletion of carbohydrates from the stem⁸³. Conversely, each plant sink events (plant height, flowering time and tillering) may increase sugar production potential in non-stress environments⁷⁵.

Drought Adaptation of Stay-Green Sorghum

Stay-green, is a post-anthesis drought resistance trait in plants that provides resistance to pre-mature leaf senescence to the plant under severe moisture stress condition during grain filling stage. It contributes to an improved yield and yield stability under moisture stress condition ¹⁰⁷.

Stay-green is an integrated droughtadaptation trait in sorghum. Delayed leaf senescence during grain filling is an emergent consequence of dynamics occurring earlier in crop growth and is largely due to an improved balance between the supply and demand of water, as well as the efficiency with which the crop converts water to biomass and grain yield^{19,51}. On the supply side, crop water use during grain filling can be enhanced by increasing water availability at anthesis or increasing water accessibility during grain filling¹¹⁸. On the demand side, crop water use can be reduced by decreasing leaf area and transpiration per unit leaf area. Leaf area can be constrained by reducing tillering⁵⁶, leaf number per culm, and individual leaf size¹⁷. Transpiration per unit leaf area can be limited by stomatal density or aperture, timing of stomatal opening, and hydraulic factors.

There are multiple ways for a plant to remain green¹⁰⁹. A stay-green phenotype may arise if the onset of senescence is delayed (type A), the rate of senescence is reduced (type B), chlorophyll is retained but photosynthesis declines (type C), greenness is retained due to rapid death at harvest (type D), or the phenotype is greener to begin with (type E). These classifications indicate that staygreen may be functional or cosmetic. Functional stay-green is characterized by the maintenance of leaf photosynthesis during grain filling (types A, B, and E)., while

cosmetic stay-green occurs when photosynthetic capacity is disconnected from leaf greenness (types C and D).

Enhanced crop productivity in waterlimited environments is dependent functional stay-green. However, not all functional stay-green is necessarily productive. For example, low sink demand relative to source, created by a small panicle or low grain number, will generate a stay-green phenotype since there is little demand for the crop to translocate carbon and nitrogen from leaves to grain^{44,95}. Therefore, selection for both staygreen and grain yield should be undertaken simultaneously in plant breeding programmes to ensure that delayed senescence is not due to low sink demand.

Stay-green improves resistance to diseases and lodging. In sorghum, genotypes with the stay-green trait continue to fill their grain generally under moisture stress conditions⁹², exhibit improved resistance to charcoal rot (*Macrophomina phaseolina*) and induced lodging¹²¹.

Genetics of Drought Resistance

The main purpose of studying the genetics of drought resistance in plants is to identify genetic factors that regulates the productivity of crops under drought stress conditions. Advances in crop improvement under water-limited conditions are only possible if drought resistance traits are identified and selected for in addition to yield ^{17,97}. Quantitative trait loci (QTLs) have been mapped ⁹⁷ on the 10 linkage groups of sorghum. They are involved in controlling traits related to yield and yield components, root systems, stay-green, plant height, flowering and maturity.

A number of traits related to drought resistance have been identified and mapped; however, the stay-green trait is recognized as the most crucial drought resistance trait in sorghum. Tuinstra *et al*¹¹¹., identified 13 genomic regions associated with post-anthesis drought tolerance in sorghum. Four QTLs were identified for yield and yield stability, seven for duration of grain development and seed weight, and two for the stay-green trait. Kebede *et al*⁵³. and Haussmann *et al*⁴².,

mentioned that there are three stay-green gene sources (B 35, SC 56 and E 36-1) from which QTLs that have been mapped onto 10 linkage groups on sorghum.

Crasta *et al*²⁶., and Xu *et al*¹²⁵., identified four stay-green QTLs and mapped two of the QTLs (*Stg1* and *Stg2*) on linkage group A, and the other two, *Stg3* and *Stg4* onto linkage group D and J. The stay-green QTLs were ranked based on their contribution to the stay-green phenotype as *Stg2*, *Stg1*, *Stg3*, and *Stg4* in their order of merit. Likewise, Xu *et al*¹²⁵., mapped three QTLs (*Chl1*, *Chl2* and *Chl3*) for chlorophyll content, and the map position coincides with the stay-green QTLs. The phenotypic association of the stay-green trait and chlorophyll content may be explained by the map position of these QTLs on the genome.

Differences in flowering time, reproductive sink strength together with variation in the environmental factors alter the expression of the stay-green trait^{41,107}. Six maturity genes (Ma1- Ma6) have been identified, and mapped onto the sorghum genome. The dominant forms of these genes cause extreme lateness⁷². Two maturity OTLs are positioned near a stay-green QTL linkage group and the major independent maturity QTLs were found to be highly correlated with stay-green rating 106,125. Tropical genotypes are found to be dominant for all four loci (Ma1-Ma4) that control the time of flowering⁸². However, substituting the dominant maturity gene, Ma1, to recessive ma1 converts a tropical sorghum to a temperate one that will flower in high latitudes⁶⁶. Tuinstra et al¹¹⁰... identified that physiological association of the maturity and stay-green trait is not well understood. The indistinct association between the two traits suggests that the earliness trait may work against reproductive sink strength during post-anthesis drought stress.

Van Oosterom *et al*¹¹⁹., found that the stay-green trait as a function of green leaf area duration (GLAD), which is affected by green leaf area at flowering, time of onset of senescence, and subsequent rate of senescence. It has been reported that the three stay-green

components appeared to be inherited independently. The inheritance of the onset of leaf senescence was additive, and the senescence rate was dominant. Consequently, GLAD was found to be partially dominant. The expression of these three factors is also affected by many environmental factors, and hence, the combined genetic effects of the three factors and the environmental factors should be considered when designing breeding programs for drought resistance 18,65. Delayed senescence in sorghum is a valuable trait that improves genotypes adaptation to drought stress, grain filling and grain yield under stress.

The expression of genes related to water deficit in plants is found to be induced by water stress, desiccation, and abscisic acid Yamaguchi-Shinozaki et observed a wide variation in the timing of induction and expression of drought related genes classifying the genes into two groups. The first group is responsible for proteins that function directly under stress tolerance, and the second group produces protein factors involved in the regulation signal transduction and gene expression under drought¹²⁶. Most of these drought-inducible genes are induced by ABA. However, various researchers have reported the existence of and ABA-dependent, ABA-independent, signal transduction cascades between the initial signal of drought stress and the expression of the genes¹⁰². Inhibition of lateral root development under moisture stress condition is reported as one mechanism of drought tolerance in plants 124. The droughtinduced inhibition of lateral root growth is partly mediated by abscisic acid. Plants that are sensitive to abscisic acid in lateral root growth are more drought tolerant than those insensitive to abscisic acid124. It was also found that abscisic acid insensitive plants have higher transpiration rates and lose water much faster than abscisic acid sensitive plants 126.

Mace *et al*⁶⁴., and Rajkumar *et al*⁸⁴., identified four QTLs for nodal root angle (*qRA1_5*, *qRA2_5*, *qRA1_8*, *qRA1_10*), three QTLs for root dry weight (*qRDW1_2*,

qRDW1_5, qRDW1_8) and eight QTLs for root volume, root fresh weight and root dry weight. Additionally, one of the root angle QTL are co-located with QTL for stay-green in sorghum and associated with grain yield⁶⁴. Recently two QTLs (qRT6 and qRT7) associated with brace roots have been mapped on sorghum Chromosome 6 and 7. Brace roots significantly contribute to effective anchorage and water and nutrient uptake during late growth and development and have a substantial influence on grain yield under water limited conditions⁵⁹. Ekanayake et al³¹., found that the inheritance of root characters was controlled equally by both additive and dominant genetic effects.

Drought Tolerance Through Breeding

The major objective of plant breeding is generating and selecting for new combinations of genes to produce genotypes with superior trait performances than those of existing genotypes, within the target environment²⁵. In any breeding programme, defining the critical traits to improve grain yield in a given target environment is critical³⁴. Identification of important traits depends on the degree of influence of a trait on yield, expression of the trait at a whole plant level, the nature of the target environment which includes, rainfall amount, distribution, onset and cessation, available soil water, nutrient status of the soil, and diseases, and economic environment. In maize, for example, it has been found that early flowering, crop water use efficiency and early vigour are important traits to breed for improve yield under drought condition⁹⁰.

The greater flexibility of sorghum in adapting to diverse climatic conditions has resulted in the evolution of tropical and temperate sorghum varieties. The tropical varieties are characterized by being tall, late harvest maturating with low photoperiod sensitivity and poor population performance. They are generally adapted to low population levels and exhibit little response to improved agricultural practices (fertilization and mechanized harvesting). The temperate sorghum varieties, on the other hand, are characterized by dwarf stems, early maturity, high yields, and less dry matter per plant⁸⁶. In the early sorghum improvement programme, conversions of tropical varieties to temperate varieties were made by substituting two dominant alleles for height and three for maturity for their recessive counterparts. The conversion programme started with hybridization of tropical and temperate varieties followed by successive backcrossing².

The most sorghum breeding programmes after the discovery of stable and heritable cytoplasm-nuclear male sterility systems in the crop is exploitation of heterosis by the production of hybrids. This discovery further enables large-scale production of commercial hybrid seed to be commercially viable²⁷. One study of the expression of hybrid vigour in grain sorghum by Doggett²⁹, revealed that there was an 84 % increase in number of seed per plant, an 82 % increase in grain weight, and a 12 % increase stover weight in the hybrids relative to the better parent.

Plant breeders have two basic approaches for breeding for drought resistance, direct and indirect breeding. Direct selection for drought is conducted under conditions where stress factors occur uniformly and predictably whereas indirect selection involves selection of genotypes under managed stress environments. However, environmental factors such as temperature and moisture are highly variable from one location to another and hence difficult to predict. Moreover, variation for stress tolerance actually exhibits a large environmental component or large genotypeby-environment interaction making direct selection for a physiological trait in a single environment difficult. As a result, indirect selection breeding is used as a preferred method where selection is made based on based on developmental traits or based on assessment of plant water status and plant function⁶⁰.

Earlier drought resistance screening was done under optimal conditions, because the maximum genetic potential of yield can only be realised under optimum conditions.

Additionally, it was believed that a high positive correlation exists between performance under optimum and stress conditions^{39,111}. However, a high genotype by environment interaction may restrict the expression of the yield potential under drought condition²⁴. Although, there is a yield penalty when selecting plants under drought condition in optimal environmental contrast to conditions.

Richards⁹⁰ and Tuinstra *et al*¹¹¹., suggested that selection under both optimal and drought conditions represents the ideal trial design to select for yield and yield stability, drought tolerance and expression of drought related traits. Hence, drought resistance and its impact on yield involve interaction between plant water relations and plant physiological functions. The interactions are further complicated by the frequency and duration of the drought, plant development stage and other stress factors such as low soil fertility and biotic stress factors.

CONCLUSION

Understanding the different drought resistance mechanisms in plants is essential when breeding for drought resistance. Stay-green is a valuable trait that improves genotype adaptation to drought stress, grain filling and grain yield under stress^{18,65}, without a yield penalty under moisture deficit conditions as compared to osmotic adjustment and early maturity¹⁷. among The balance characters maintains adequate productivity by providing a spectrum of effective drought tolerance mechanisms. An early maturing genotype yields less compared to a late maturing genotype in a favourable environment. This is because drought escape by shortening the growing period is made at the expense of the crops genetic yield potential.

REFERENCES

Ackerson, R.C. and Hebert, R.R.,
 Osmoregulation in cotton in response to
 water stress. I. Alterations in
 photosynthesis, leaf conductance,

- translocation, and ultrastructure. *Plant Physiol.*, **67:** 484-488 (1981).
- 2. Acquaah, G., Principles of plant genetics and breeding. Blackwell Publishing, Carlton, Australia (2007).
- 3. Amede, T. and Schubert, S., Mechanism of drought resistance in grain legumes I: Osmotic adjustment. *Ethiop. J. Sci.*, **26**: 37-46 (2003).
- 4. Amelework, B., Shimelis, H., Tongoona, P. and Laing, M., Physiological mechanisms of drought tolerance in sorghum, genetic basis and breeding methods: A review. *Afr. J. Agric. Res.*, **10(31):** 3029-3040 (2015).
- 5. Baalbaki, R.Z., Zurayk, R.A., Bleik, M.M. and Saxena, N.S., Germination and seedling development of drought tolerant and susceptible wheat under moisture stress. *Seed Sci. Technol.*, **27**: 291-302 (1999).
- 6. Bawazir, A.A. and Idle, D.B., Drought resistance and root morphology in sorghum. *Plant Soil.*, **119:** 217-221 (1989).
- 7. Bittman, S. and Simpson, G.M., Drought effect on leaf conductance and leaf rolling in forage grasses. *Crop Sci.*, **29:** 338-334 (1989).
- 8. Blum, A. and Ebercon, A., Genotypic responses in sorghum to drought stress. III. Free proline accumulation and drought resistance. *Crop Sci.*, **16:** 428-431 (1976).
- 9. Blum, A. and Sullivan, C.Y., The comparative drought resistance of landraces of sorghum and millet from dry and humid regions. *Ann. Bot.*, **57:** 835-846 (1986).
- Blum, A., (2004). Sorghum Physiology.
 pp. 141-223. *In*: Nguyen, H.T., and A.
 Blum (eds.) Physiology and biotechnology integration for plant breeding. Marcel Dekker Inc., NY.
- Blum, A., Arkin, G.F. and Jordan, W.R., Sorghum root morphogenesis and growth.
 I. Effect of maturity genes. *Crop Sci.*, 17: 149-153 (1977).

- 12. Blum, A., Crop responses to drought and the interpretation of adaptation. Plant Growth Regul., 20: 135-148 (1996).
- 13. Blum, A., Drought resistance and its improvement. In: Blum A. (ed). Plant Breeding for Water-Limited Environments. Springer Science Business Media, NY, pp. 53-137 (2011).
- 14. Blum, A., Drought resistance, water-use efficiency, and yield potential-are they compatible, dissonant, or mutually exclusive? Aust. J. Agric. Res., 56: 1159-1168 (2005).
- 15. Blum, A., Golan, G., Mayer, J. and Sinmena, B., The effect of dwarfing genes on sorghum grain filling from remobilized stem reserves, under stress. Field Crops Res., **52:** 43-54 (1997).
- 16. Blum, A., Sinmena, B., Mayer, J., Golan, and Shpiler, L., Stem reserve mobilization supports wheat grain filling under heat stress. Aust. J. Plant Physiol., **21:** 771-781 (1994).
- 17. Borrell, A. K., Hammer, G. L., Douglas, A. C. L., Does maintaining green leaf area in sorghum improve yield under drought? I. Leaf growth and senescence. Crop Science, 40: 1026-1037 (2000a).
- 18. Borrell, A.K., Hammer, G.L. and Henzell, R.G., Does maintaining green leaf area in sorghum improve yield under drought? II. Dry matter production and yield. Crop Sci., 40: 1037-1048 (2000b).
- 19. Borrell, A.K., Jordan, D.R., George-Jaeggli, B., Hammer, G.L., Van Oosterom, E., Klein, P. and Mullet, J., Fine-mapping candidate genes for 'stay-green' sorghum: are we there yet? International Conference on Integrated Approaches to Improve Crop Production Drought-Prone **Environments** under (InterDrought-III).. Shanghai, China: Shanghai Academy of Agricultural Science. L 5.03 (2009).
- 20. Borrell, A., Jordan, D., Mullet, J., Henzell, B. and Hammer, G., Drought adaptation in sorghum. In: Ribaut J.M. (ed) Drought adaptation in cereals. The Haworth Press

- (2006).21. Camacho, B.S.E., Hall, A.E. and Kufman, M.R., Efficiency and regulation of water
- transport in some woody and herbaceous species. Plant Physiol., 54: 169-172 (1974).
- 22. Campos, H., Cooper, M., Habben, J.E., Edmeades, G.O. and Schussler, J.R., Improving drought tolerance in maize: A view from industry. Field Crops Res., 90: 19-34 (2004).
- 23. Ceccarelli, S., Grando, S., Baum, M. and Udupa, S.M., Breeding for drought resistance in a changing climate. In: Rao S., Ryan J. (eds) Challenges and Strategies of Dryland Agriculture. CSSA Special Publication no. 32, CSSA and ASA, Madison, WI, pp. 167-190 (2004).
- 24. Chapman, S.C., Cooper, M., Butler, D.G. Henzell. R.G., Genotype environment interactions affecting grain sorghum: I Characteristics that confound interpretation of hybrid yield. Aust. J. Agric. Res., 51: 197-207 (2000a).
- 25. Chapman, S., Cooper, M., Podlich, D. and Hammer, G., Evaluating plant breeding strategies by simulating gene action and dryland environment effects. Agron. J., **95:** 99-113 (2003).
- 26. Crasta, O.R., Xu, W.W., Rosenow, D.T., Mullet, L. and Nguyen, H., Mapping of post-flowering drought resistance traits in grain sorghum: Association between QTLs influences premature senescence and maturity. Mol. Gen. Genet., 262: 579-588 (1999).
- 27. Dar, W.D., Reddy, B.V.S., Gowda, C.L.L. and Ramesh, S., Genetic resources of **ICRISAT-mandated** enhancement crops. Curr. Sci., 91: 880-884 (2006).
- 28. Dingkuhn, M., Audebert, A.Y., Jones, M.P., Etienne, K. and Sow, A., Control of stomatal conductance and leaf rolling in O. sativa and O. glaberrima upland rice. Field Crops Res., 61: 223-236 (1999).
- 29. Doggett, H., Sorghum. 2nd ed. Longman Scientific and Technical, London (1988).

- 30. Egilla, J.N., Davies, E.T. and Drew, J.M.C., Effect of potassium on drought resistance of Hibiscus rosa-sinensis cv. Leprechaun: Plant growth, leaf macro- and micronutrient content and root longevity. Plant Soil., 229: 213-224 (2001).
- 31. Ekanayake, I.J., O'Toole, J.C., Garrity, D.P. and Masajo, T.M., Inheritance of root characters and their relations to drought resistance in rice. Crop Sci., 25: 927-933 (1985).
- 32. Esau, K., Plant Anatomy (2nd Ed.). Wiley, New York (1965)
- 33. Farooq, M., Wahid, A., Kobayashi, N., Fujita, D. and Basra, S.M.A., Plant drought stress: effects, mechanisms and management. Agron Sustain Dev., 29: 185-212 (2009).
- 34. Fernandez, G.C.J., Effective selection criteria for assessing stress tolerance. In: Kuo CG (ed). Proceedings of the International Symposium on "Adaptation of Vegetables and Other Food Crops in Temperature and Water Stress". 13-16 Aug 1991. Tainan, Taiwan. pp. 257-270 (1992).
- 35. Flower, D.J., Rani, A.U. and Peacock, J.M., Influence of osmotic adjustment on the growth, stomatal conductance and light interception of contrasting sorghum lines in a harsh environment. Aust. J. Plant Physiol., 17: 91-105 (1990).
- 36. Fortmeier, R. and Schubert, S., Salt tolerance of maize (Zea mays L.).: The role of sodium exclusion. Plant Cell Environ., 18: 1041-1047 (1995).
- 37. Gollan, T., Passioura, J.B. and Munns, R., Soil water status affects the conductance of fully turgid wheat and sunflower leaves. Aust. J. Plant Physiol., 13: 459-464 (1986).
- 38. Grote, E.M., Ejeta, G. and Rhodes, D., Inheritance of glycinebetaine deficiency in sorghum. Crop Sci., 34: 1217-1220 (1994).
- 39. Habyarimana, E., Laureti, D., De Ninno, M. and Lorenzoni, C., Performance of biomass sorghum (Sorghum bicolor L. Moench) under different water regimes in

- Mediterranean region. Indus. Crops Prod., **20:** 23-28 (2004).
- 40. Hammer, G.L., Farquhar, G.D. and Broad, I.J., On the extent of genetic variation for transpiration efficiency in sorghum. Australian J. Agric. Res., 48: 649-655 (1997).
- 41. Harris, K., Subudhi, P.K., Borrell, A., Jordan, D., Rosenow, D., Nguyen, H., Klein, P., Klein, R. and Mullet, J., Sorghum stay-green QTL individually reduce post-flowering drought induced leaf senescence. J. Exp. Bot., 58: 327-338 (2007).
- 42. Haussmann, B.I.G., Mahalakshmi, V., Reddy, B.V.S., Seetharama, N., Hash, C. T. and Geiger, H. H., QTL mapping of stay-green in two sorghum recombinant inbred populations. Theor. Appl. Genet., **106:** 133-142 (2002).
- 43. Heckathorn, S.A. and E.H., Effect of leaf rolling on gas exchange and leaf temperature of Andropogon gerardii and Spartina pectinata. Botanical Gazette, **152:** 263-268 (1991).
- 44. Henzell, R.G. and Gillieron, W., Effects of partial and complete panicle removal on the rate of death of some Sorghum bicolor under moisture genotypes stress. Queensland J. Agri. Animal Sci., 30: 291-299 (1973).
- 45. HongBo, S., ZongSuo, L. and MingAn, S., Osmotic regulation of 10 wheat (Triticum aestivum L.) genotypes at soil water deficits. Colloids Surf. B. Biointerfaces, **47:** 132-139 (2006).
- 46. Hsiao, T.C., O'Toole, J.C., Yambao, E.B. and Turner, N.C., Influence of osmotic adjustment on leaf rolling and tissue death in rice (Oryza sativa L.). Plant Physiol., **75:** 338-341 (1984).
- 47. Impa, S.M., Nadaradjan, S., Boominathan, P., Shashidhar, G., Bindumadhava, H. and Sheshshayee, M.S., Carbon isotope discrimination accurately reflects variability in WUE measured at a whole plant level in rice. Crop Sci., 45: 2517-2522 (2005).

- 48. Inoue, T., Inanaga, S., Sugimoto, Y. and Siddig, K.E., Contribution of pre-anthesis assimilates and current photosynthesis to grain yield, and their relationships to drought resistance in wheat cultivars grown under different soil moisture. *Photosynthetica*, **42**: 99-104 (2004).
- 49. Izanloo, A., Condon, A.G., Langridge, P., Tester, M. and Schnurbusch, T., Different mechanisms of adaptation to cyclic water stress in two South Australian bread wheat cultivars. *J. Exp. Bot.*, **59:** 3327-3346 (2008).
- 50. Jones, M.M. and Rawson, H.M., Influence of rate of development of leaf water deficit upon photosynthesis, leaf conductance, water use efficiency and osmotic potential in sorghum. *Physiol. Plant.*, **45:** 103-111 (1979).
- 51. Jordan, D.R., Hunt, C.H., Cruickshank, A.W., Borrell, A.K., Henzell, R.G., The relationship between the stay-green trait and grain yield in elite sorghum hybrids grown in a range of environments. *Crop* Science, **52:** 1153–1161 (2012).
- 52. Kadioglu, A. and Terzi, R., A dehydration avoidance mechanism: Leaf rolling. *Bot. Rev.*, **73**: 290-302 (2007).
- 53. Kebede, H., Subudhi, P.K., Rosenow, D.T. and Nguyen, H.T., Quantitative trait loci influencing drought tolerance in grain sorghum (*Sorghum bicolour L. Moench*).. *Theor. Appl. Genet.*, **103:** 266-276 (2001).
- 54. Khan, H.R., Link, W., Hocking, T.J. and Stoddard, F.L., Evaluation of physiological traits for improving drought tolerance in faba bean (*Vicia faba* L.).. *Plant Soil.*, **292:** 205-217 (2007).
- 55. Khanna-Chopra, R. and Sinha, S.K., Enhancement of drought induced senescence by the reproductive sink in fertile lines of wheat and sorghum. *Ann. Bot.*, **61:** 649-653 (1988).
- 56. Kim, H.K., Luquet, D., Van Oosterom, E., Dingkuhn, M. and Hammer, G., Regulation of tillering in sorghum: genotypic effects. *Annals of Botany*, **106**: 69–78 (2010).

- 57. Lambrides, C.J., Chapman, S.C. and Shorter, R., Genetic variation for carbon isotope discrimination in sunflower: Association with transpiration efficiency and evidence for cytoplasmic inheritance. *Crop Sci.*, **44:** 1642-1653 (2004).
- 58. Levitt, J., Responses of Plants to Environmental Stress, Water, Radiation, Salt and Other Stresses. Academic Press, NY (1980).
- Li, R., Han, Y., Lv, P., Du, R. and Liu, G., Molecular mapping of the brace root traits in sorghum (Sorghum bicolor L. Moench).. Breed. Sci., 64: 193-198 (2014).
- 60. Ludlow, M.M. and Muchow, R.C., A critical evaluation of traits for improving crop yield in water-limited environments. *Advan. Agron.*, **43:** 107-153 (1990).
- 61. Ludlow, M.M., Adaptive significance of stomatal response to water stress, pp. 123-138. In N. C. Turner and P- J. Kramer, (eds.) Adaptation to water and high temperature stress. Wiley Interscience; New York (1980).
- 62. Ludlow, M.M., Fisher, M.J. and Wilson, J.R., Stomatal adjustment to water deficits in three tropical grasses and tropical legume grown in controlled conditions and in the field. *Aust. J. Plant Physiol.*, **12**: 131-149 (1985).
- 63. Ludlow, M.M., Santamaria, J.M. and Fukai, S., Contribution of osmotic adjustment to grain yield in *Sorghum bicolor* L. Moench under water-limited conditions II: Water stress after anthesis. *Australian J. Agric. Res.*, **41**: 67-78 (1990).
- 64. Mace, E., Singh, V., Van Oosterom, E., Hammer, G., Hunt, C. and Jordan, D., QTL for nodal root angle in sorghum (*Sorghum bicolor* L. Moench). co-locate with QTL for traits associated with drought adaptation. *Theor. Appl. Genet.*, **124:** 97-109 (2012).
- 65. Mahalakshmi, V. and Bidinger, F.R., Evaluation of stay-green sorghum germplasm lines at ICRISAT. *Crop Sci.*, **42:** 965-974 (2002).

- 66. Major, D.J., Rood, S.B. and Miller, F.R., Temperature and photo period effects mediated by the sorghum maturity genes. *Crop Sci.*, **30**: 305-310 (1990).
- 67. Matsuura, A., Inanaga, S. and Sugimoto, Y., Mechanism of interspecific differences among four graminaceous crops in growth response to soil drying. *Japanese J. Crop Sci.*, **65**: 352-360 (1996).
- 68. McGuire, S.J., Farmer management of sorghum diversity in Eastern Ethiopia, In: Almekinders, CJM, de Boef, WS (eds.)., Encourage diversity: The conservation and development of plant genetic resources. Intermediate Technoloy Publication, London, pp. 43-48 (2000).
- 69. Merah, O., Deleens, E., Souyris, I., Nachit, M. and Monneveux, P., Stability of carbon isotope discrimination and grain yield in durum wheat. *Crop Sci.*, **41**: 677-681 (2001).
- 70. Meyer, W.S. and Ritchie, J.T., Resistance to water flow in the Sorghum plant. *Plant Physiol.*, **65:** 33-39 (1980).
- 71. Mitra, J., Genetics and genetic improvement of drought resistance in crop plants. *Curr. Sci.*, **80:** 758-763 (2001).
- 72. Morgan, P.W., Finlayson, S.A., Childs, K.L., Mullet, J.E. and Rooney, W.L., Opportunities to improve adaptability and yield in grasses: Lessons from sorghum. *Crop Sci.*, **42:** 1781-1799 (2002).
- 73. Mortlock, M.Y. and Hammer, G.L., Genotype and water limitation effects on transpiration efficiency in sorghum. *J. Crop Prod.*, **2:** 265-286 (1999).
- 74. Moussa, H.R. and Abdel-Aziz, S.M., Comparative response of drought tolerant and drought sensitive maize genotypes to water stress. *Aust. J. Crop Sci.*, **1:** 31-36 (2008).
- 75. Murray, S.C., Sharma, A., Rooney, W.L., Klein, P.E., Mullet, J.E., Mitchell, S.E. and Kresovich, S., Genetic improvement of sorghum as a bio-fuel feedstock: I. QTL for stem sugar and grain non-structural carbohydrates. *Crop Sci.*, **48:** 2165-2179 (2008).

- 76. Nour, A.M., Weibel, D.E. and Tood, G.W., Evaluation of root characteristics in grain sorghum. *Agron. J.*, **70:** 217-218 (1978).
- 77. Pace, P.F., Cralle, H.T., El-Halawaney, S.H.M., Cothern, J.T. and Sensemen, S.A., Drought induced changes in shoot and root growth of young cotton plants. *J. Cotton. Sci.*, **3**: 183-187 (1999).
- 78. Passioura, J.B., Research on drought and salinity: Avenues for improvements. *Aust. J. Plant Physiol.*, **13:** 191-201 (1986).
- 79. Plaut, Z., Carmi, A. and Grava, A., Cotton root and shoot response to subsurface drip irrigation and partial wetting of the upper soil profile. *Irrig. Sci.*, **16:** 107-113 (1996).
- 80. Prasad, P.V.V., Pisipati, S.R., Mutava, R.N. and Tuinstra, M.R., Sensitivity of grain sorghum to high temperature stress during reproductive development. *Crop Sci.*, **48:** 1911-1917 (2008).
- 81. Prince, A.H., Cairns, J.E., Horton, P., Jones, H.G. and Griffiths, H., Linking drought-resistance mechanisms to drought avoidance in upland rice using a QTL approach: Progress and new opportunities to integrate stomatal and mesophyll responses. *J. Exp. Bot.*, **53**: 1569-1576 (2002).
- 82. Quinby, J.R., Sorghum Improvement and the Genetics of Growth. Texas A and M University Press, College Station, TX. (1974).
- 83. Rajewski, J.F. and Francis, C.A., Defoliation effects on grain fill, stalk rot, and lodging of grain sorghum. *Crop Sci.*, **31:** 353-359 (1991).
- 84. Rajkumar, B.F., Kavil, S.P., Girma, Y., Arun, S.S., Dadakhalandar, D., Gurusiddesh, B.H., Patil, A.M., Thudi, M., Bhairappanavar, S.B., Narayana, Y.D., Krishnaraj, P.U., Khadi, B.M. and Kamatar, M.Y., Molecular mapping of genomic regions harbouring QTLs for root and yield traits in sorghum (Sorghum bicolor L. Moench).. Physiol. Mol. Biol. Plants., 19: 409-419 (2013).

- 85. Ramu, S.V., Palaniappan, S. and Panchanathan, R., Growth and dry matter partitioning of sorghum under moisture stress condition. *J. Agron. Crop Sci.*, **166**: 273-277 (2008).
- 86. Rao, N.G.P., Murty, U.R. and Rana, B.S., Sorghum. pp. 213-238. In: Chapra V.L., Prakash S. (eds) Evolution and Adaptation of Cereal Crops. Science Publishers Inc. Enfield, USA (2002).
- 87. Rebetzke, G.J., Condon, A.G., Richards, R.A. and Farquhar, G.D., Selection for reduced carbon isotope discrimination increases aerial biomass and grain yield of rainfed bread wheat. *Crop Sci.*, **42**: 739-745 (2002).
- 88. Redmannm, R.E., Adaptation of grasses to water stress: Leaf rolling and stomate distribution. *Ann. Missouri Bot. Gard.*, **72**: 833-842 (1985).
- 89. Richards, R.A. and Passioura, J.B., Seminal root morphology and water use of Wheat. II. Genetic variation. *Crop Sci.*, **21:** 253-255 (1981).
- 90. Richards, R.A., Defining selection criteria to improve yield under drought. *Plant Growth Regul.*, **20:** 157-166 (1996).
- 91. Robertson, M.J., Fukai, S., Ludlow, M.M. and Hammer, G.L., Water extraction by grain sorghum in a sub-humid environment. II. Extraction in relation to root growth. *Field Crop Res.*, **33**: 99-112 (1993).
- 92. Rosenow, D.T. and Clark, L.E., Drought tolerance in sorghum. p. 18-31. In: Loden H.D., Wilkinson D. (eds). Proceeding of 36th Annual Corn and Sorghum Industry Research Conference. Chicago, IL. 9–11 Dec. 1981. American Seed Trade Association, Washington, DC (1981).
- 93. Rosenow, D.T. and Clark, L.E., Drought and lodging resistance for a quality sorghum crop. In: Proceedings of the 5th annual corn and sorghum industry research conference (Chicago, IL, 6-7 December 1995)., American Seed Trade Association, Chicago, IL, pp 82-97 (1995).
- 94. Rosenow, D.T., Ejeta, G., Clark, L.E., Gilbert, M.L., Henzell, R.G., Borrell, A.K.

- and Muchow, R.C., Breeding for pre- and post-flowering drought stress resistance in sorghum. In: Rosenow, D.T, Yohe, J.M (eds) Proceedings of the international conference on genetic improvement of sorghum and pearl millet (Lubbock, TX, 22–27 September 1996)., ICRISAT, Lubbock, India, pp 400-411 (1996).
- 95. Rosenow, D.T., Quisenberry, J.E., Wendt, C.W. and Clark, L.E., Drought tolerant sorghum and cotton germplasm. *Agricultural Water Management*, **7:** 207–222 (1983).
- 96. Routley, R., Broad, I., McLean, G., Whish, J. and Hammer, G., The effect of row configuration on yield reliability in grain sorghum: I. Yield, water use efficiency and soil water extraction. Proceeding of the Eleventh Australian Agronomy Conference. Geelong, Australia (2003).
- 97. Sanchez, A.C., Subudhi, P.K., Rosenow, D.T. and Nguyen, H.T., Mapping QTLs associated with drought resistance in sorghum (*Sorghum bicolor L. Moench*). *Plant Mol. Biol.*, **48:** 713-726 (2002).
- 98. Shangguan, Z.P., Lei, T.W., Shao, M.A. and Xue, Q.W., Effects of phosphorous nutrient on the hydraulic conductivity of sorghum (*Sorghum vulgare* Pers) seedling roots under water deficiency. *J. Integr. Plant Biol.*, **47:** 421-427 (2005).
- 99. Shao, H.B., Liang, Z.S. and Shao, M.A., Osmotic regulation of 10 wheat (*Triticum aestivum* L.). genotypes at soil water deficits. *Colloids Surfaces B: Biointerfaces.*, **47:** 132-139 (2006a).
- 100. Shao, H.B., Chen, X.Y., Chu, L.Y., Zhao, X.N., Wu, G., Yuan, Y.B., Zhao, C.X. and Hu, Z.M., Investigation on the relationship of proline with wheat anti-drought under soil water deficits. *Colloids Surfaces B: Biointerfaces.*, **53:** 113-119 (2006b).
- 101. Shao, H.B., Liang, Z.S., Shao, M.A. and Wang, B.C., Changes of anti-oxidative enzymes and membrane peroxidation for soil water deficits among 10 wheat genotypes at seedling stage. *Colloids*

- Krupa et al
 Int. J. Pure App. Biosci. 5 (4): 221-237 (2017)

 Surfaces B: Biointerfaces., 42: 107-113
 tolerance developmen

 (2005).
 developmen
- 102. Shinozaki, K. and Yamaguchi-Shinozaki, K., Molecular responses to dehydration and low temperature: Differences and cross-talk between two stress signalling pathways. *Curr. Opin. Plant Biol.*, **3:** 217-223 (2000).
- 103. Simpson, G.M., (1981). Water Stress in Plants. Praeger, NY.
- 104. Sivaramakrishnan, S., Patell, V.Z., Flower, D.J. and Peacock, J.M., Proline accumulation and nitrate reductase activity in contrasting sorghum lines during midseason drought stress. *Physiologia Plantarum.*, **74:** 418-426 (2006).
- 105. Stiller, W.N., Read, J.J., Constable, G.A. and Reid, P.E., Selection for water use efficiency traits in a cotton breeding program: cultivar differences. *Crop Sci.*, **45:** 1107-1113 (2005).
- 106. Taiz, L. and Zeiger, E., Stress physiology. pp. 671–681. *In*: Taiz L, Zeiger E, (eds.) Plant Physiology, 4th ed. Sinauer Associates, Inc., Sunderland, MA (2006).
- 107. Tao, Y.Z., Henzell, R.G., Jordan, D.R., Butler, D.G., Kelly, A.M. and McIntyre, C.L., Identification of genomic regions associated with stay-green in sorghum by testing RILs in multiple environments. *Theor. Appl. Genet.*, **100**: 1225-1232 (2000).
- 108. Teulat, B., Merah, O., Sirault, X., Borries, C., Waugh, R. and This, D., QTLs for grain carbon isotope discrimination in field-grown barley. *Theor. Appl. Genet.*, **106:** 118-126 (2002).
- 109. Thomas, H. and Howarth, C.J., Five ways to stay green. *J. Exp. Bot.*, **51**: 329-337 (2000).
- 110. Tuinstra, M.R., Ejeta, G. and Goldsbrough, P., Evaluation of nearly-isogenic sorghum lines contrasting for QTL markers associated with drought tolerance. *Crop Sci.*, **38:** 835-842 (1998).
- 111. Tuinstra, M.R., Grote, E.M., Goldbrough, P.M. and Ejeta, G., Genetic analysis of post-flowering drought

- 221-237 (2017) ISSN: 2320 7051 tolerance and components of grain development in *Sorghum bicolor* L. Moench. *Mol. Breed.*, **3:** 439-448 (1997).
- Tumwesigye, E.K. and Musiitwa, F., 112. Characterizing drought patterns appropriate development and transfer of drought resistance maize cultivar in Uganda. . In: 7th Eastern and Southern Africa Regional Maize Conference and Symposium on Low-Nitrogen Drought Tolerance in Maize. 11th-15th February, 2002. Nairobi, Kenya. CIMMYT-Kenya and Kenya Agricultural Research Institute (KARI)., Nairobi. pp. 260-262 (2002).
- 113. Turner, N.C, Schulze, E.D. and Gollan, T., The response of stomata and leaf gas exchange to vapour pressure deficits and soil water content. II. In the mesophytic herbaceous species *Helianthus annuus*. *Oecologia*, **65**: 348-355 (1985).
- 114. Turner, N.C., Begg, J.E. and Tonnet, M.L., Osmotic adjustment of sorghum and sunflower crops in response to water deficits and its influence on the water potential at which stomata close. *Aust. J. Plant physiol.*, **5:** 597-608 (1978).
- 115. Turner, N.C., Crop water deficits: A decade of progress. *Adv. Agron.*, **39:** 1-48 (1986).
- 116. Twomlow, S., Mugabe, F.T., Mwale, M., Delve, R., Nanja, D., Carberry, P. and Howden, M., Building adaptive capacity to cope with increasing vulnerability due to climatic change in Africa A new approach. *Phy. Chem. Earth.*, **33:** 780-787 (2008).
- 117. Vajrabhaya, M., Kumpun, W. and Chadchawan, S., The solute accumulation: The mechanism for drought tolerance in RD23 rice (*Oryza sativa* L.). lines. *Sci. Asia.*, **27**: 93-97 (2001).
- 118. Van Oosterom, E.J., Borrell, A.K., Deifel, K.S. and Hammer, G.L., Does increased leaf appearance rate enhance adaptation to postanthesis drought stress in sorghum? *Crop Science*, **51**: 2728–2740 (2011).

- 119. Van Oosterom, E.J., Jayachandran, R. and Bidinger, F.R., Diallel analysis of the stay-green trait and its components in sorghum. *Crop Sci.*, **36:** 549-555 (1996).
- 120. Wood, A.J. and Goldsbrough, P.B., Characterization and expression of dehydrins in water-stresses *Sorghum bicolor*. *Physiologia Plantarum*., **99:** 144-152 (1997).
- 121. Woodfin, C.A., Rosenow, D.T. and Clark, L.E., Association between the stay-green trait and lodging resistance in sorghum. Agronomy Abstracts. ASA, Madison, WI (1988).
- 122. Wright, G.C., Smith, R.C.G. and Mcwilliams, J.R., Differences between two grain sorghum genotypes in adaptation to drought stress. Crop growth and yield responses. *Aust. J. Agric. Res.*, **34:** 615-626 (1983).
- 123. Xin, Z., Aiken, R. and Burke, J., Genetic diversity of transpiration efficiency in sorghum. Field *Crops Res.*, **111:** 74-80 (2008).
- 124. Xiong, L., Wang, R., Mao, G. and Koezan, J., Identification of drought tolerance determinants by genetic analysis

- of root responses to drought stress and abscisic acid. *Plant Physiol.*, **142:** 1065-1074 (2006).
- 125. Xu, W., Subudhi, P.K., Crasta, O.R., Rosenow, D.T., Mullet, J.E., Nguyen, H.T., Molecular mapping of QTLs conferring stay-green in sorghum (*Sorghum bicolor* L. Meonch). *Genome*, **43:** 461-469 (2000).
- 126. Yamaguchi-Shinozaki, K., Kasuga, M., Liu, Q., Nakashima, K., Sakuma, Y., Abe, H., Shinwari, Z.K., Seki, M. and Shinozaki, K., Biological mechanisms of drought stress response. Japan International Research Centre for Agricultural Sciences (JIRCAS). Working Report 23, Tsukuba, Japan, pp. 1-8 (2002).
- 127. Yang, W J., Rich, P.J., Axtell, J.D., Wood, K.V., Bonham, C.C., Ejeta, G., Mickelbart, M.V. and Rhodes, D., Genotypic variation for glycinebetaine in sorghum. *Crop Sci.*, 43: 162-169 (2003).
- 128. Younesi, O. and Moradi, A., The effect of water limitation in the field on sorghum seed germination and vigour. *Aust. J. Basic Appl. Sci.*, **3:** 1156-1159 (2009).